

## Note

# Characterization of the polysaccharide antigen of *Klebsiella pneumoniae* O:9 lipopolysaccharide

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The lipopolysaccharides (LPS) of *Klebsiella pneumoniae* have been implicated as virulence determinants. Although some 72 K-serospecific capsular antigens have been described for *Klebsiella* sp., it has been suggested that there exist only 8 associated unique LPS O-polysaccharide antigens in this gram-negative bacterial species<sup>1</sup>.

In recent re-investigations of *Klebsiella* LPS O-chains<sup>2,3</sup>, originally proposed structures have required revision, and the production of multiple unique O-chain structures by a given designated *Klebsiella* serotype has been demonstrated<sup>2,3</sup>. The present investigation of the *Klebsiella* O:9 LPS polysaccharide has shown that it is a polymer of a branched pentasaccharide unit composed of only D-galactose residues. The newly deduced structure of the O-antigen differs from the originally proposed structure<sup>4</sup> in the mode of linkage at its branch point and, as a consequence, also in the structure of the backbone D-galactan polymer.

The O-polysaccharide, extracted by the method of Johnson and Perry<sup>5</sup> as described in the Experimental section, had  $[\alpha]_D +93^\circ$  (*c* 1.5, H<sub>2</sub>O) and, by quantitative GLC<sup>6</sup> and capillary GLC of the derived (*R*)-2-butyl glycosides<sup>7</sup>, was shown to be composed of D-galactose (90%). Anal. Found: C, 39.77; H, 5.71; N, 0.20; and ash 0%. The <sup>1</sup>H NMR spectrum of the native O-chain showed signals at  $\delta$  2.12 and 2.15 indicative of *O*-acetyl methyl protons while its <sup>13</sup>C NMR spectrum (Fig. 1B) also showed *O*-acetyl signals at  $\delta$  21.20 and 21.18 (CH<sub>3</sub>CO) and 173.5 and 174.7 (CH<sub>3</sub>CO). The <sup>1</sup>H NMR spectrum of the *O*-deacetylated O-polysaccharide (dil NH<sub>4</sub>OH) showed *inter alia* four H-1 signals, at  $\delta$  5.31 (1 H, *J*<sub>1,2</sub> 3 Hz), 5.25 (2 H, *J*<sub>1,2</sub> 0.5 Hz), 5.08 (1 H, *J*<sub>1,2</sub> 3 Hz), and 5.05 (1 H, *J*<sub>1,2</sub> 3 Hz). The <sup>13</sup>C NMR spectrum of the same *O*-deacetylated O-polysaccharide (Fig. 1A) showed

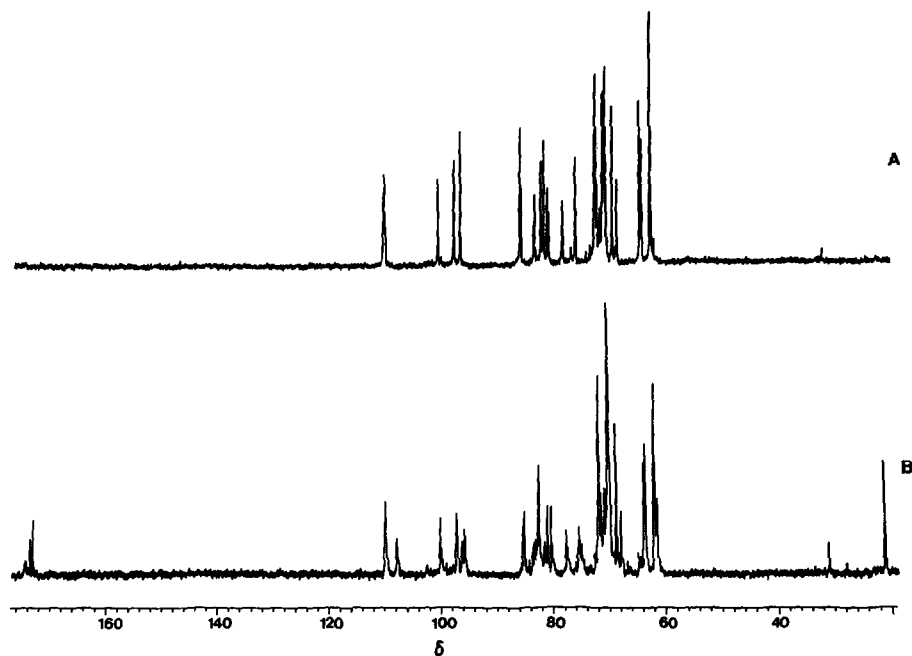


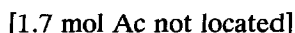
Fig. 1.  $^{13}\text{C}$  NMR spectra (125 MHz,  $47^\circ\text{C}$ ) of: A, O-Deacetylated *K. pneumoniae* O:9 LPS O-polysaccharide and B, native *K. pneumoniae* O:9 LPS O-polysaccharide.

*inter alia* five C-1 signals, at  $\delta$  110.2 ( $J_{\text{C,H}}$  176 Hz), 110.1 ( $J_{\text{C,H}}$  176 Hz), 100.3 ( $J_{\text{C,H}}$  170 Hz), 97.6 ( $J_{\text{C,H}}$  172 Hz), and 96.3 ( $J_{\text{C,H}}$  171 Hz). These chemical shift and  $J_{\text{C,H}}$  coupling data were indicative of two  $\beta$ -D-Galf and three  $\alpha$ -D-Galp residues in a repeating pentasaccharide unit of the D-galactan polymer. The O-acetyl content of the native O-chain corresponded to  $\sim 1.7$  mol per repeating unit.

The O-deacetylated O-chain had  $[\alpha]_{\text{D}} + 86^\circ$  ( $c$  5.4,  $\text{H}_2\text{O}$ ). Anal. Found: C, 40.90; H, 5.64; N, 0.66; and ash 0%. On methylation analysis<sup>8</sup> with the aid of GLC-MS the alditol acetates of 2,3,4,6-tetra-O-methyl-D-galactose ( $t_{\text{R}}$  1.08), 2,5,6-tri-O-methyl-D-galactose ( $t_{\text{R}}$  1.36), 2,4,6-tri-O-methyl-D-galactose ( $t_{\text{R}}$  1.47) and 4,6-di-O-methyl-D-galactose ( $t_{\text{R}}$  1.82) were identified in the molar ratio 1:2:1:1. This analysis is consistent with the proposed pentasaccharide structure and the presence in the repeating unit of a nonreducing D-Galp end-group, two  $\rightarrow 3$ -D-Galf residues, a  $\rightarrow 3$ -D-Galp residue, and a branched, di-O-substituted residue,  $\rightarrow 2,3$ -D-Galp.

As expected from the methylation evidence, the periodate oxidation of the O-deacetylated O-chain resulted in the oxidation of the D-Galp nonreducing end-group and the exocyclic C-6–C-5 diol systems of the D-Galf residues. The product of the Smith type<sup>9</sup> mild hydrolysis of the reduced ( $\text{NaBH}_4$ ), periodate-oxidized O-chain afforded, on Sephadex G-50 chromatography, a polysaccharide, eluting at the void volume of the system, which had  $[\alpha]_{\text{D}} + 62^\circ$  ( $c$  1.1,  $\text{H}_2\text{O}$ ) and

The identification of 4,6-di-*O*-methyl-D-galactose in the methylation analysis of the O-chain, considered in conjunction with the characterization of the O-chain linear backbone and the  $\alpha$ -D configurations of the D-Galp residues, indicates that the nonreducing  $\alpha$ -D-Galp end-group is (1  $\rightarrow$  2)-linked to  $\alpha$ -D-Galp units in the backbone structure. From the NMR spectra of the O-chain it can be deduced that this substitution follows a regular pattern consistent with the O-chain being composed of a repeating pentasaccharide unit in which the D-Galf and D-Galp residues have the  $\beta$  and  $\alpha$  configurations respectively. Thus, the structure is as shown here.



The location and proportion of the *O*-acetyl substituents appears to be variable and dependent upon bacterial growth conditions, but may however play a role in the serology of the O:9 antigen. In the <sup>13</sup>C NMR spectra of native O-chain preparations the presence of extra C-1 signals from the D-Galf residues at  $\delta \sim 108$  (Fig. 1B) suggests that a significant proportion of O-acetylation was always present at the 2 position of these units.

Cells of *Klebsiella pneumoniae* O9:K<sup>-</sup> (NRCC 4378, CWK 48, from strain 121205), grown in 3.7% (w/v) brain–heart infusion (Difco) at 37°C in a Microfirm fermenter, were extracted by the hot aqueous phenol method<sup>5</sup>, and subsequent isolation procedures were performed as previously described<sup>2</sup>. Ultracentrifugation afforded LPS from the aqueous phase (6% yield) and the phenol phase (0.9% yield). Fission of the aqueous phase LPS with hot 2% acetic acid (2 h, 100°C) gave

an insoluble lipid A (~ 10%) and Sephadex G-50 chromatography of the water soluble products gave the O-polysaccharide (69% yield) eluting at the void volume of the system. Glycan hydrolyses, aldose identifications, periodate oxidations, methylation analyses, and  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy were also done under the same conditions as previously described<sup>2</sup>.

GLC–MS analysis of acetylated methyl alditols was done with an OV-17 fused silica capillary column using a temperature program from 200°C (2 min) to 240°C at 1°C/min. Retention times are quoted relative to 1,5-di-O-acetyl-2,3,4,6-tetra-O-methyl-D-glucitol ( $t_{\text{R}} = 1.00$ ).  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts ( $\delta$ ), quoted relative to tetramethylsilane, were measured from internal acetone ( $\delta_{\text{H}}$  2.225,  $\delta_{\text{C}}$  31.07); coupling constants are given in Hertz.

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